

A NOTE ON SALT ACCUMULATION WITH ECOPHYSIOLOGICAL COMMENTS

DEVENDRA SINGH, SUNEETA DADORIA, R M AGARWAL*, MADHUP SHRIVASTAVA,
MOHAMMAD ABASS AHANGER

School of Studies in Botany, Jiwaji University, Gwalior, MP India 474011
agarwalrm@rediffmail.com

(Revised 10th Jan 2014; Accepted 22nd February,2014)

ABSTRACT

Increased industrialization and urbanization has resulted in conversion of arable land into marginal land. A few grasses in particular *Vetiveria zizanioides* L and *Sporobolus ioclados* L can withstand adverse environmental conditions because of their higher tolerance level. Present study was aimed to assess the uptake and partitioning of different ions by these two grasses i.e. *Vetiveria zizanioides* L and *Sporobolus ioclados* L. Accumulation of different ions has been noticed and the content of ion accumulated increased with the developmental stage without having too much impact on their growth and morphology.

Keywords : *Vetiveria zizanioides* L, *Sporobolus ioclados* L, sodium, potassium, calcium and chloride.

Introduction

Vetiveria zizanioides and *Sporobolus ioclados* are grasses belonging to family Poaceae and are commonly known as “Khus” and “Sawarri” respectively. *Vetiveria zizanioides* is characterised by having short rhizomes, deep, extensive and penetrating root system. Stem of *Vetiveria zizanioides* L. is tall and leaves are long, thin and rigid (Erskine, 1992; Troung, 1999; Hellin and Haigh, 2002; Ke *et al.*, 2003). These plants have a strong adaptability to adverse conditions, which make them ideal for soil, water conservation and erosion control in arid and semi arid regions. Their greater adaptability is attributed to fast growth, easy planting, high survival rate, also never turn into a kind of weed (Lavania, 2000).

Vetiveria zizanioides and *Sporobolus ioclados* have been reported to withstand drought, long periods of inundation, extreme temperatures, soil acidity and alkalinity

(pH from 3.3 to 10.5) (Erskine, 1992; Dalton *et al.*, 1996; Truong and Loch, 2004; Zhou and Yu, 2009). Unique morphological, physiological and ecological characteristics render these plant species capable of growing in harsh environmental and soil conditions. *Vetiveria zizanioides* and *Sporobolus ioclados* accumulate toxic heavy metals mainly in roots and least in shoots (Yang *et al.*, 2003). *Vetiveria zizanioides* and *Sporobolus ioclados* have been reported to possess high tolerance to metals like Al, Mn and heavy metals like Cd, Cr, N, Pb, Hg, Se, Zn and metalloids such as arsenic present in the soils (Troung, 2000; Cheng *et al.*, 2004; Chiu *et al.*, 2005). This tolerance to the heavy metals by grasses is often attributed to their capability to accumulate metals in above ground tissues without affecting the root and shoot growth. Moreover, mycorrhizal association within the roots of *Vetiveria zizanioides* make it sturdy enough to withstand high metal concentration

in the soil (Roongtanakiat and Chairaj, 2002; Cheng *et al.*, 2004). Salinity is one of the most deleterious abiotic stresses, which adversely influences plant growth, development and crop productivity in both irrigated and non-irrigated areas of the world (Jungklang *et al.*, 2003; Ashraf *et al.*, 2008; Zhu, 2008). Growth of plants is generally reduced due to osmotic stress induced by the accumulation of sodium and chloride (Munns and Termaat, 1986; Zhou and Yu, 2009).

High concentration of salts in the soil solution also interferes with balanced absorption of essential nutritional ions by plants (Tester and Devenport, 2003). Salt tolerant plants provide good material for investigating the adaptation mechanisms (Ashraf, 2003). Soil salinity is an important feature of landscape of arid countries especially where artificial irrigation is practiced (Flower, 2004). Halophytic plants have the capability to minimize the detrimental effects by morphological means and physiological or biochemical processes such as osmotic adjustment achieved by accumulating osmolytes like K^+ and proline (Jacoby, 1999). Focus has been on using tolerant plant species for conservation of soil and water along with phytoremediation of heavy metals and other pollutants from contaminated fields (Pang *et al.* 2003; Lai and Chen 2004).

Height of most grasses is adversely affected at the higher levels of salinity which might be due to less availability of water and toxicity of sodium chloride (Munns, 2002). However, seeds of *Sporobolus ioclados* germinated in NaCl upto 500mM (Gulzar *et al.*, 2003) and shoot length of *Vetiveria zizanioides* was increased by 18.60% at 200 mM NaCl concentration but was adversely affected at higher (300mM) NaCl concentration. So it appears that the grass species exhibit salinity tolerance upto 200 mM NaCl as far as growth in terms of shoot length is concerned (Mane *et al.*, 2011). *Vetiveria zizanioides* has been reported to function as an effective adsorbent for the removal of

fluoride from aqueous solution (Puthenveedu *et al.*, 2012). It was with this viewpoint that accumulation of salts in different parts of *Vetiveria zizanioides* and *Sporobolus ioclados* at different developmental stages was evaluated for the present work.

Material and Methods

Young plants of *Vetiveria zizanioides* and *Sporobolus ioclados* were collected from Malanpur (Bhind) and Jiwaji University campus and planted in pots at Botanical Garden of Jiwaji University, Gwalior. Different plant parts were collected at two different stages after plantation i.e. 20 and 40 days after plantation. These parts were washed with distilled water and dried in oven at 60°C for 2 days. Dried plant parts were used for analysis.

Estimation of K, Na and Ca.

Estimation of K, Na and Ca was done flame photometrically as adopted by Tomar and Agarwal (2013). One gram of dried plant material was digested using triacid mixture ($H_2SO_4 + HNO_3 + HClO_4$ in 9:3:1 ratio). After digestion final volume was made up to 100 ml using distilled water and filtered. Colourless filtrate was directly read on flame photometer for estimation of Na, K and Ca using separate filters and calculations were done using standard curves.

Estimation of chloride.

Estimation of chloride was done following method of Eaton *et al.* (2005). One gram dry plant sample was boiled in 100 ml distilled water on a water bath for 30 minutes. After cooling the extract was filtered and 25 ml of this filtrate was used for analysis. Few drops (5-6) of 5% potassium chromate (indicator) were added and titrated against 0.1 N $AgNO_3$ solution till a permanent brick red precipitate appears.

Results and Discussion

An increase in uptake and accumulation of Potassium, sodium, calcium and chloride contents was found at later stage of plantation. Leaves of both *Sporobolus ioclados* L and *Vetiveria zizanioides* L partitioned greater amounts of potassium and calcium than roots. However, sodium accumulated to a greater degree in roots than leaves indicating greater tolerance of roots to sodium (Tables 1 and 2). Salt tolerance is a complex process which involves adaptations at physiological and genetic levels in both more tolerant (halophytes) and less tolerant plants (Flower, 2004). Synthesis and accumulation of compatible organic osmolytes such as polyamines, glycine betaine, proline and free sugars is an important adaptation of plants to mitigate stress (Bartels and Sunkar, 2005). Salinity affects plant growth through the osmotic and ionic effects thereby disrupting the integrity of cellular membranes, uptake of essential nutrients, function of photosynthetic apparatus and many other physiological and biochemical processes (Zhu, 2008).

Osmotic adjustment is considered to be an important component of salt tolerance mechanisms in plants and is usually defined as a decrease in the cell sap osmotic potential resulting due to net increase in intracellular osmolytes to prevent the loss of cell water. Intracellular osmolytes include inorganic ions normally absorbed from medium and organic solutes which are synthesized and transported within the plant (Carvajal, *et al.*, 1999; Bajji *et al.*, 2001; Annick *et al.*, 2004; Navarro *et al.*, 2007). Inorganic ions, mainly include K^+ , Na^+ , Ca^{2+} , Mg^{2+} , Cl^- , NO_3^- , SO_4^{2-} , $H_2PO_3^-$ which however, can cause ionic toxicity when accumulated in higher concentrations, particularly Na^+ and Cl^- (Martínez *et al.*, 2003; Ottow *et al.*, 2005; Touchette, 2007; Yang *et al.*, 2007).

Potassium is an important macro element which improves plant growth and resistance

against environmental stresses. Potassium is involved in several physiological processes like stomatal regulation, osmoregulation, enzyme activation, maintaining energy status, charge balance, photosynthesis, protein synthesis and translocation (Beringer and Trolldenier 1978; Marschner 1995). Excess rain or irrigation can cause leaching of potassium through the soil. Potassium deficiency results in reduced growth, short internodes, scorched leaf margins, necrosis, reduced lateral breaks and increases susceptibility to wilting. Studies have shown that the application of K fertilizer mitigates the adverse effects of drought on plant growth (Andersen *et al.*, 1992; Studer, 1993; Sangakkara *et al.*, 2001).

Sodium can substitute for potassium and K deficiency symptoms of many plants are reduced by Na (Hylton *et al.*, 1967; Amin and Joham, 1968). Na has been shown to be an essential element for Bermuda grassland for certain C_4 plants (Brownell and Crossland, 1972). Sodium requirement was first demonstrated for the bladder srat-bush, a perennial pasture species of arid inland areas of Australia have since then surveyed 32 species of plants for their sodium requirement. The metabolic basis for the sodium requirement may be related to the transport of pyruvate, a critical intermediate in the C_4 pathway between the bundle-sheath and mesophyll cells (Brownell and Wood, 1957; Brownell and Crossland, 1972).

Sodium has specific function in concentration of CO_2 in C_4 plants. Many halophytic plants are able to take advantage of similarity between sodium and potassium and have adapted to grow in areas of high salt whereas growth of other less adapted plants is limited due to high salinity (Greenway and Munns, 1980; Glenn *et al.*, 1999). Uptake and movement under natural conditions is possible to increase the cycling of sodium through plants by suitable nutrient management practices and at times sodium containing wastes could be used to

Table 1: Percent potassium, sodium, calcium and chloride in *Sporobolus ioclados* L.

Plant part	Potassium		Sodium		Calcium		Chloride	
	20 days after plantation	40 days after plantation	20 days after plantation	40 days after plantation	20 days after plantation	40 days after plantation	20 days after plantation	40 days after plantation
Roots	0.043±0.003	0.1652±0.015	0.0250±0.002	0.1381±0.027	0.025±0.013	0.250±0.00	0.0011±0.00	0.0142±0.00
Stem	0.085±0.006	0.4415±0.002	0.0378±0.002	0.0846±0.001	0.2175±0.017	0.410±0.021	0.0017±0.00	0.0237±0.001
Leaves	0.122±0.002	0.5417±0.007	0.0463±0.001	0.0985±0.009	0.254±0.013	0.571±0.00	0.0042±0.00	0.0385±0.004
Flower	0.0729±0.008	0.3164±0.002	0.0306±0.003	0.0829±0.001	0.2785±0.005	0.431±0.021	0.0025±0.00	0.0397±0.004

Table 2: Percent potassium, sodium, calcium and chloride in *Vetiveria zizanioides* L.

Plant part	Potassium		Sodium		Calcium		Chloride	
	20 days after plantation	40 days after plantation	20 days after plantation	40 days after plantation	20 days after plantation	40 days after plantation	20 days after plantation	40 days after plantation
Leaves	0.870±0.0012	0.605±0.0029	0.165±0.0078	0.108±0.008	0.146±0.0020	1.053±0.001	0.025±0.0000	0.038±0.0028
Roots	0.279±0.0023	0.657±0.0202	0.221±0.0640	0.234±0.0053	0.120±0.0500	1.169±0.010	0.009±0.0012	0.034±0.0000

grow plants, which could then provide food, O₂ and clean water for the crew (Wheeler *et al.*, 2002).

Present study reveals greater uptake and accumulation of ions with the time of plantation and their partitioning is more in the upper plant parts i.e. leaves and

stem. Increase in content of ions with the developmental stage reflects about the enhanced potential of *Sporobolus ioclados* L. and *Vetiveria zizanioides* L. to accumulate salts which can be quite useful in conversion of marginal land into arable land. Obviously, further studies should be rewarding.

REFERENCES

- Amin, J.V. and Joham, H.E. (1968). The cations of the cotton plant in sodium substituted potassium efficiency. *Soil Sci.* **105**: 248-254.
- Andersen, M. N., Jensen, C.R., Losch, R. (1992). The interaction effects of potassium and drought in field-grown barley., Yield, water-use efficiency and growth. *Acta Agric. Scand. Sect. B Soil Plant Sci*, B: 34-44.
- Annick, M.M. Elisabeth, P. and Gerard, T. (2004). Osmotic Adjustment, Gas Exchanges and Chlorophyll Fluorescence of a Hexaploid Triticale and Its Parental Species under Salt Stress, *J. Plant Physiol.* vol. **161**: 25-33.
- Ashraf, M., (2003). Relationships between leaf gas exchange characteristics and growth of differently adapted populations of Blue panicgrass (*Panicum antidotale* Retz.) under salinity or waterlogging. *Plant Sci.* **165**: 69-75
- Ashraf, M., Athar, H.R., Harris P.J.C. and Kwon T.R. (2008). Some prospective strategies for improving crop salt tolerance. *Adv Agron.*, **97**: 45-110.
- Bajji, M., Lutts, S. and Kinet, J.M. (2001). Water Deficit Effects on Solute Contribution to Osmotic Adjustment as a Function of Leaf Ageing in Three Durum Wheat cultivars Performing Differently in Arid Conditions, *Plant Sci.* vol. **160**: 669-681.
- Bartels, D. and Sunkar, R. (2005). Drought and salt tolerance in plants. *Criti. Rev. Plant Sci.* **24**: 23-58.
- Beringer, H. and Trolldenier, G. (1978). Influence of K nutrition on the response to environmental stress, in: Potassium Research - Review and Trends. Proceedings of the 11th Congress of the International Potash Institute. Internat. Potash Inst. Bern. 189-222.
- Brownell, P.F. and Crossland. C. J. (1972). The requirement for sodium as a micronutrient by species having the C₄ dicarboxylic photosynthetic pathway. *Plant Physiol.* **49**: 794-797.
- Carvajal, M., Martinez, V. and Alcaraz, C.F. (1999). Physiological Function of Water-Channels, as Affected by Salinity in Roots of Paprika Pepper, *Physiol. Plant.*, vol. **105**: 95-101.

- Cheng, H., Yang, X., Liu, A., Fu, H. and Wan, M. (2004). A study on the performance and mechanism of soil-reinforcement by herb root system. In: Proceedings of third international vetiver conference, Guangzhou, China. 384–390.
- Chiu, K.K., Ye, Z.H. and Wong, M.H. (2005). Growth of *Vetiveria zizanioides* and *Phragmites australis* on Pb/Zn and Cu mine tailings amended with manure compost and sewage sludge: A greenhouse study. *Bioresour. Technol.* **97**: 158-170.
- Dalton, P.A., Smith, R.J. and Truong, P.N.V. (1996). Vetiver grass hedges for erosion control on a cropped flood plain: hedge hydraulics. *Agr. Water Manage* **31**: 91–104.
- Eaton, A., Clesceri, L., Rice, E. and Greenberg, A. (2005). Standard Methods for the Examination of Water and Wastewater 21st edition, American Public Health Association, American Water Works Association and Water Environmental Federation, Washington.
- Erskine, J.M. (1992). Vetiver grass: its potential use in soil and moisture conservation in Southern Africa. *S Afr J Sci* **88**: 298–299.
- Flower, T.J. (2004). Salt tolerance is complex genetically and physiologically. *J. Exp. Bot.* **55**: 307-319.
- Glenn, E.P., Brown, J.J. and Blumwald, E. (1999). Salt tolerance and crop potential of halophytes. *Crit. Rev. Plant Sci.* **18**: 227–255.
- Greenway, H. and Munns, R. (1980). Mechanisms of salt tolerance in non halophytes. *Annu., Rev. Plant Physiol. Plant Mol. Biol.* **31**: 149–190.
- Gulzar, S., Khan, M. A. and Ungar, I. A. (2003). Salt tolerance of a coastal salt marsh grass. *Communications in Soil Science and Plant Analysis.* **34**: 2595-2605.
- Hellin, J. and Haigh, M. J. (2002). Better land husbandry in Honduras, towards the new paradigm in conserving soil, water and productivity. *Land Deg. Dev.* **13**(3): 233–250.
- Hylton, L.O., Ulrich, A. and Cornelius, D.R. (1967). Potassium and sodium interrelations in growth and mineral content of Italian ryegrass. *Agron. J.* **59**: 311-315.
- Jacoby, B. (1999). Mechanism involved in salt tolerance of plants. In: *Handbook of Plant and Crop Stress*, ed. M., Pessaraki, Marcel Dekker, Inc. New York. 97–124.
- Jungklang, J., Usui, K. and Matsumoto, H. (2003). Differences in Physiological Responses to NaCl Between Salt-Tolerant (*Sesbania rostrata* Brem. and Oberm.) and Non-Tolerant (*Phaseolus vulgaris* L.). *Weed Biol Manage.* **3**: 21–27.
- Ke, C., Feng, Z., Wu, X. and Tu, F. (2003). Design principles and engineering samples of applying vetiver eco-engineering technology for steep slope and riverbank stabilization. In: *Proc 3rd Int'l Conf. on Vetiver*. Guangzhou, China. China Agricultural Press, Beijing. pp. 365–374.
- Lai, H.Y. and Chen, Z.S. (2004). Effect of EDTA on solubility of cadmium, zinc and lead and their uptake by rainbow pink and vetiver grass. *Chemosph.* **55**: 421–430.
- Lavania, U.C. (2000). Primary and secondary centers of origin of vetiver and its dispersion., In: Chomchalow, N., Barang, N. (Eds.) Proceedings of the 11th International Conference on Vetiver: Vetiver and Environment. Office of Royal Development Project Board, Bangkok, Thailand. 424–427.
- Mane, A.V. (2011). Emir, J. *food Agricu.* **23**(1): 59-70.
- Marschner, H. (1995). Why can sodium replace potassium in plants? In: *Proceedings of 8th Colloquium of International Institute of Potash Institute.* 50–63.
- Munns, R. (2002). Comparative physiology of salt tolerance and water stress. *Plant Cell and Environment* **25**: 239-250
- Munns, R. and Termaat, A. (1986). Whole-plant responses to salinity. *Australian Journal of Plant Physiology* **13**: 143–160.
- Navarro, A., Ban, N. S., Olmos, E. and Sánchez-Blanco, M. J. (2007). Effects of Sodium Chloride on Water Potential Components, Hydraulic Conductivity, Gas Exchange and Leaf Ultrastructure of *Arbutus unedo* Plants, *Plant Sci.* **17**: 2473–2480.
- Ottow, E.A., Brinker, M., Teichmann, T., Fritz, E., Kaiser, W., Brosché, M., Kangasjärvi, J., Jiang, X.N. and Polle, A. (2005) *Populus euphratica* Displays Apoplastic Sodium Accumulation, Osmotic Adjustment by Decreases in Calcium and Soluble Carbohydrates, and Develops Leaf Succulence under Salt Stress, *Plant Physiol.* **vol. 139**: 1762–1772.
- Pang, J., Chan, G.S.Y., Zhang, J., Liang, J. and Wong, H.M. (2003). Physiological aspects of vetiver grass for rehabilitation in abandoned metalliferous mine waste. *Chemosphere* **52**: 1559–1570.
- Puthenveedu, Sadasivan, Pillai, Harikumar, Chonattu, Jaseela, Tharayil and Megha. (2012). Water Quality Division, Centre for Water Resources Development and Management, Kozhikode, India. **4**: 245-251.
- Roongtanakiat, N. and Chairaj, P. (2002). Vetiver grass for remediating soil contaminated with heavy metals. *Proc. 17th World Congr. Soil Sci.*, Bangkok, Thailand.
- Sangakkara, U.R., Frehner, M. and Nosberger, J. (2001). Influence of soil moisture and fertilizer potassium on the vegetative growth of mungbean (*Vigna radiata* L. Wilczek) and cowpea (*Vigna unguiculata* L. Walp). *J. Agron. Crop Sci.* **186**: 73–81.
- Studer, C. (1993). Interactive effects of N, P, K, nutrition and water stress on the development of young maize plants. Ph.D. Thesis, ETHZ, Zurich, Switzerland.

- Tester, M. and Davenport, R. (2003). Sodium tolerance and sodium transport in higher plants, *Ann. Bot.* **91**: 503–507.
- Tomar, N. S. and Agarwal, R. M. (2013). Influence of Treatment of *Jatropha curcas* L. Leachates and Potassium on Growth and Phytochemical Constituents of Wheat (*Triticum aestivum* L.) *American Journal of Plant Sciences.* **4**: 1134-1150.
- Touchette, B.W. (2007). Seagrass-Salinity Interactions: Physiological Mechanisms Used by Submersed Marine Angiosperms for a Life at Sea, *J. Exp. Mar. Biol. Ecol.* vol. **350**: 194–215.
- Truong, P. and Loch, R. (2004). Vetiver System for erosion and sediment control. In: Proc. 13th International soil conservation organization conference, Brisbane, Australia, 1-6.
- Truong, P.N. (1999). Vetiver grass technology for land stabilization, erosion and sediment control in the Asia Pacific region. In: Proc. First Asia Pacific Conference on Ground and Water Bioengineering for Erosion Control and Slope Stabilization, Manila, Philippines. Int Eros Control Assoc, Steamboat Springs, USA. 72–84.
- Truong, P.N.V. (2000). The global impact of Vetiver grass technology on the protection. In: The 2nd Int. Vetiver Conf., Thailand.
- Yang, C.W., Chong, J.N., Li, C.Y., Kim, C.M., Shi, D.C. and Wang, D.L. (2007). Osmotic Adjustment and Ion Balance Traits of an Alkali Resistant Halophyte *Kochia sieversiana* during adaptation to Salt and Alkali Conditions, *Plant Soil*, vol. **294**: 263–276.
- Zhou, Q. and Yu, B.J. (2009). Published in *Fiziologiya Rastenii*, Vol. **56(5)**: 751–758.
- Zhu, J.K. (2008). Plant salt tolerance. *Trends Plant Sci.* **6**: 66–71.