

Pharmaceutical Sciences, 2022, 28(1), 15-26 doi:10.34172/PS.2021.36 https://ps.tbzmed.ac.ir/

Review Article



Development of Imaging and Liquid Biomarker Analysis for Breast Cancer Screening: A Review

Atmedi Surendra^{1,6}, Tina Rostinawati¹, Riezki Amalia²

¹Department of Pharmaceutical Biology, Faculty of Pharmacy, Padjadjaran University, Sumedang, Indonesia. ²Department of Pharmacology and Clinical Pharmacy, Faculty of Pharmacy, Padjajaran University, Sumedang, Indonesia.

Article Info

Article History: Received: 2 January 2021 Accepted: 18 June 2021 ePublished: 8 July 2021

Keywords:

- -Biomarker -Breast cancer -Detection
- -Diagnostic
- -Imaging
- -Screening

Abstract

Background: Breast cancer screening tests could reduce the mortality rates for breast cancer patients. Screening and detection are the keystone of cancer prevention and may significantly minimize the death rates in breast cancer patients for long-term. In this review, we would like to present a comprehensive summary from recent publications of the current development for breast cancer screening, classification of breast cancer based on pathological diagnosis, as well as development of breast cancer detection.

Methods: The sources of the articles were collected from research published in the PubMed, NCBI databases and manual searches without time restriction based on review of the title, abstract and full review of the articles, using the keywords "breast cancer", "diagnostic", "screening", "imaging", "biomarker" and the combination of these terms. The criteria excluded in selecting references were articles that are not written in English, newspapers, and posters.

Results: Of the 146 articles that were selected, there were 103 articles included. Breast cancer screening consists of imaging and pathological assessment such as invasive biopsies of tumor tissue and measurement of biomarkers. The recent development of breast cancer screening utilizing different models and methods like biomarkers were being reviewed. For imaging methods, there are mammography, digital breast tomosynthesis (3D mammography), magnetic resonance imaging (MRI), and ultrasonography. For pathological assessment, there are primary biomarker analysis for breast cancer (estrogen receptor, progesterone receptor, HER2, KI67 index) and liquid biomarker analysis from blood or saliva samples. Additionally, there are some diagnostic kit models for breast cancer screening that were in use such as NanoString nCounter[®], MammaTyper[®], CellSearch System[™], and AdnaTest BreastCancer[™].

Conclusion: Each of these methods has its own limitations. Therefore, the development of breast cancer models should be more sensitive, reliable, approachable and less harmful.

Introduction

Breast cancer is one of the highest prevalent cancers in which the incidence rate is higher in developed countries.¹ There are some risk factors such as age, hormone status, family history, genetic predisposition, environment, lifestyle, and population structure that could alter the prevalence of breast cancer which is different every person in many regions.² Furthermore, the molecular patterns in primary and metastases tumors of patients are different.³

Early breast cancer screening is a keystone of cancer prevention and could reduce the mortality rates.⁴ The main problems are the lack of community access such as source availability caused by excessively high costs, lack of knowledge, hours of operation, or distance from the access source.⁵ Highly sensitive, rapid, reliable, and accessible early-stage breast cancer diagnostic is important factor to decrease mortality rates and improve breast cancer detection quality by reducing recalls of false positive results and unnecessary biopsies.⁶

Breast cancer clinical examinations consist of imaging diagnosis and pathological assessment such as invasive biopsies of tumor tissue and measurement of biomarkers.⁷ Imaging techniques contain mammography and ultrasound detection for the breast tissues as a target. Pathological assessment should be conducted based on biomarker detection techniques like core needle biopsy.⁸ This paper reviews the current development for early-stage breast cancer screening, classification of breast cancer based on the patient's pathological diagnosis results, and development of breast cancer detection to overcome emerging problems such as availability, accessibility, and patients with certain condition.

*Corresponding Author: Atmedi Surendra, E-mail: atmedisurendra@yahoo.com

©2021 The Author(s). This is an open access article and applies the Creative Commons Attribution License (http://creativecommons.org/licenses/by-nc/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, as long as the original authors and source are cited.

Methods

Search Strategy

The materials for this study were selected from PubMed, NCBI databases and manual search without time restriction based on review of the title, abstract and full review of the articles. The search keyword included: breast cancer, diagnostic, screening, imaging, biomarker and the combination of these terms. The excluded criteria in selecting references were articles that are not written in English, newspapers, and posters.

By using keywords that have been specified, 146 articles were obtained. Of the 146 articles, 103 articles were selected for evaluating the recent development breast cancer screening using different models and methods.

Results

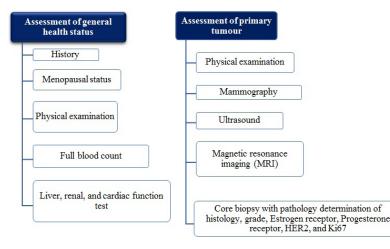
As there no cure yet for breast cancer, there are prevention methods for women to reduce the risk of breast cancer.9 The primary prevention method is by utilizing prophylactic surgery or chemoprevention for woman with high risk.9 The secondary prevention method is by utilizing early detections such as clinical breast examination and screening methods like imaging and biomarker analysis that offer the most effective, practical, and viable methods for women around the world.9 The main objectives of early breast cancer screening are to enable women to be able to undergo less invasive diagnostics and to identify asymptomatic cancer that leads to ideal outcomes before the breast cancer progresses through both physical breast examinations (e.g., mammographic imaging) and pathological assessment as breast cancer screening modalities.10 The next assessments required are personal medical history, family history, renal and liver function tests, calcium and alkaline phosphatase levels, another physical examination, determining menopausal status and a full blood count.8 The pathological results consist of the histological type, breast cancer grade (TNM stage), immunohistochemical (IHC) evaluation of breast cancer biomarker such as estrogen receptor (ER) and progesterone receptor (PgR), and human epidermal growth factor receptor (HER2) gene expression status (Figure 1).¹¹

Imaging

In most cases, breast cancer is detected by screening test or through analyzing symptoms that occur on patients (e.g., pain or a palpable mass) and associated with the detection of tumor size, metastasis, and would likely require chemotherapy to reduce morbidity and improve the patient's survival rates.¹² Breast magnetic resonance imaging (MRI) or ultrasonography may be considered as additional screenings for high-risk woman.¹⁰ These imaging techniques (Table 1) have advantages for avoiding unnecessary breast biopsies. The early diagnostic method accuracy was determined by sensitivity and specificity from final assessment that were defined as true positive (TP), true negative (TN), false negative (FN), and false positive (FP). The calculation formula for sensitivity is = $(TP/(TP + FN) \times 100 \text{ and for specificity is} = TN/(TN + FP)$ $\times 100^{13}$

Mammography and Digital Breast Tomosynthesis (3D mammography)

Mammography is the basic method of breast cancer diagnostic that only has average sensitivity and could reduce inpatient with high dense breasts because of overlying breast parenchyma or lesions from overlapping tissues.⁴ Mammography also has disadvantages such as the use of ionizing radiation, relatively high false-output rates, and providing uncomfortable examination for patients.¹⁴ The impact of such disadvantages is that the patients must be recalled for another exact assessment and require improvement for them to be viable for breast cancer screening.¹⁵ There are digital breast tomosynthesis (DBT) and contrast-enhanced digital mammography to improve the limitations of the conventional mammography like specificity.¹⁶ Contrast-enhanced mammography the techniques have the potential to encourage good initial results with 85.2% sensitivity and 66.1% specificity.17 Contrast-enhanced mammography could reduce radiation exposure, health care costs, and false-positive rates.¹⁸ The development of digital mammography creates digital breast tomosynthesis that could provide analysis of





3D mammographic data that presents high detail and answering some disadvantages from overlapping tumor tissue.¹⁹ Digital mammography may have drawbacks that reduces sensitivity because fibroglandular tissue may be overlying tumors.¹⁰ Digital breast tomosynthesis using X-ray kit that could move over a limited arc angle and reconstruct the tissues in thin slices to reduce overlapping tumor tissue.¹⁶ The addition of digital breast tomosynthesis (DBT) may improve the detection rate of breast cancer and reduce patient recalls rate.²⁰ One concern on using this method for breast cancer screening is the digital breast tomosynthesis contains double radiation dose over conventional digital mammography alone.¹²

Magnetic Resonance Imaging (MRI) and Ultrasonography

Both magnetic resonance imaging (MRI) and ultrasonography are decent tools for evaluating and diagnosing abnormalities of breast tissues especially for specific populations such as very high-risk women with mutations of BRCA1 and BRCA2 or women with dense breast.¹⁰

Magnetic resonance imaging (MRI) displays multiple cross-sections image by involving magnetic field. The resolution of magnetic resonance imaging (MRI) could be increased by applying contrast agent⁴ and already been recommended for detecting breast cancer in high risk breast cancer patients.²¹ Magnetic resonance imaging (MRI) is less specific but more sensitive compared to

Table 1. Imaging methods for early breast cancer screening.

mammography or ultrasound for detecting small tumors in patients with high-risk.²² Several studies recommend combination of mammography and magnetic resonance imaging (MRI) for women with high breast cancer risk. The benefit is demonstrated by comparing a group of women with BRCA1 and BRCA2 mutations.²³

Breast ultrasonography is a good method as it is widely available, cost effective, and could improve examination sensitivity through the detection of the breast cancer mass shape.²⁴ Some observational and clinical studies have shown that the combination of ultrasonography and mammography could increase the detection rates and sensitivity of breast cancer screening in women which have dense breasts.²⁵ Breast ultrasonography has already been introduced as an additional screening for high-risk patients.²⁶ However, there are some disadvantages of breast ultrasonography such as possible failures in screening many tumors due to the similar features of cancerous and normal tissues. Moreover, this method requires experienced radiologists and could affect the specificity and sensitivity of the results.⁴

Biomarker Analysis

There are certain indications of cancer progression called biomarkers (Table 2) that could be expressed in/on tumor tissues such as biomolecules from microRNAs, some mutated genes, and cell surface receptor proteins.²⁷ As a diagnostic tool, biomarkers assemblies demand such non-

Туре	Use	Sensitivity*	Specificity*	Advantage	Disadvantage	Ref.
Mammography	Mass detection. Displays bone, blood vessel and soft tissue image.	67.8%	75%	for early breast cancer screening, proven to	Radiation exposure, relative- ly high false-positive rates and false-negative rates, not suitable for women (patients) with high risk such as dense breasts, and slightly uncomfortable assessment.	4, 12
Ultrasonography	Detects the mass shape of breast cancer at an early stage.	83%	34%	stage breast cancer in	Requires experienced operator during examination and displays image with low resolution.	4, 24
Magnetic Resonance Imaging (MRI)	Displays small details images of soft tissues.		26.4%	Screening for patient with high risk especially for young women or patients with dense breasts.	Overestimation of tumor size, expensive method.	4, 12,22
Contrast- enhanced Mammography	Detects area that shows vascularizati- on in patient.		66.1%	Improving sensitivity for conventional mammo- graphy, better lesion representation in dense breast where the image may be blocked by fibroglandular tissue	evidence for breast cancer	17
Digital Breast Tomosynthesis	Examines actual breast lesions, allows better separation in tumorous and normal tissues illustration of lesions.		96.49	May improve lesion detection and reduces false positive and recall rates.	Radiation dose approxi- mately is twice that of mammography.	6, 19, 20

* Breast composition and the types of cancer could affect sensitivity and specificity of the methods.

invasive techniques and should be differently identified in healthy individuals.28 The serum could be collected as common analytes for a biomarker as expressed on the cell surface extra cellular domains (ECD).²⁷ Breast cancer biomarkers have two classifications: overexpressed biomolecules-based biomarkers and stage-dependent biomarkers.²⁹ Biomarkers may contain genetic sequencing information for some individuals that have BRCA mutation.7 Basic expression techniques for biomarkers are enzyme linked immunosorbent assay (ELISA), immunohistochemistry (IHC), and radioimmunoassay.³⁰ Core biopsy with following immunohistochemical breast cancer molecular subtypes evaluation is currently the basic method for breast tissue assessment as it has relatively high sensitivity compared to other methods.³¹ Immunohistochemical examination is utilized to facilitate the classification of breast cancer subtypes.³² The main molecular biomarkers that is related to breast cancer are progesterone receptor (PR), estrogen receptor (ER), Mib1/ Ki-67 proliferation index and human epidermal growth factor receptor (HER2) as they are remarkably established in the standard care of breast cancer patients.³³ ER and PR could stimulate the growth of breast epithelium, play an important role as sex steroid receptors and could express around 75% of all breast cancers. Additionally, poor prognosis could also be related to overexpression of HER2.³⁴ (Table 2).

Estrogen Receptor (ER) and Progesterone Receptor (PR)

Sexual hormones usually provide an impact to growth of breast cancer tissue. Estrogen receptor is one of the most notable biomarkers because estrogen receptor acts as transcription factors that promote survival, proliferation, and invasion of the cell.³³ Estrogen receptor is ligandregulated. The DNA-binding domains are the main components of estrogen receptor which specifically binds with high affinity on estrogen response elements (ERE sequence) and organizes the transcription rates of ligandbinding domain.35 There are ERa and ERB which are the two forms of estrogen receptor that are differentially expressed in tissues. Both ERa and ERB manage cell differentiation and proliferation by binding estradiol in the normal mammary gland.³⁶ ER-positive patient could reduce recurrence and mortality from breast cancer by using ER as the target therapy and endocrine therapy (tamoxifen and aromatase inhibitors) as the treatment.³² Nowadays, immunohistochemistry is the standard practice evaluation of estrogen-progesterone receptor expression.37 There are guidelines that establish the inspection criteria and proficiency testing for hormone receptor to increase its accuracy. The specimens of breast resection must be arranged as quickly as possible (within 1 hour from resection) in a fixative with adequate volume.³⁸

Human Epidermal Growth Factor Receptor 2 (HER2)

The human epidermal growth factor receptor 2 (HER2) genes are localized on chromosome 17 and they are

regularly expressed at low levels in all epithelial cells. HER2 are one of the significant components for cancer survival and proliferation.³³ High levels expression of mRNA and protein product by HER2 genes amplification could conduct self-sufficiency and oncogenic resultant signaling in growth signals, continuous angiogenesis, uncontrolled growth, and amplify metastasis processes that could encourage carcinogenesis.³² The total results of HER2 that are amplified in patients range approximately from 15– 30% of breast cancers cases.³⁹

Immunohistochemistry for testing HER2 protein overexpression has been developed, and may become the standard procedure for detecting invasive breast carcinomas.⁴⁰ The amplification of HER2 gene could be analyzed by fluorescence (FISH), silver-enhanced (SISH) or chromogenic in situ hybridization (CISH) and directly linked to mRNA and protein expression levels that could be analyzed through ELISA test, Western blot, immunohistochemistry, real time PCR or Northern blot.³³ Immunohistochemistry has already been assessed as the standard test in determining the HER2 status, which has advantages such as quicker results, the ability to display morphological tumor appearance, and the ability to maintain stained preserved tissues to degrade slower over time.³²

MiB1 / Ki67

Mib1/Ki-67 is a biomarker measured through proliferation index as the parameters for predictive and prognostic markers. In most cases, breast cancer patients experience worse outcomes as they are expressing high levels of Ki67.³² Mib1/Ki-67 index decreases for patients who are provided with post-treatment of neoadjuvant therapies. It becomes a decent predictor for improved clinical outcomes. However, the ASCO guidelines have not included Mib1/Ki67 index as a primary assessed marker for breast cancer prognosis due to the lack of standardization of testing and the interpretation of this index.³³

BRCA

The BRCA genes comprehend a group of tumor suppressor genes.⁴¹ Patients with BRCA mutation carriers could increase their lifetime risk of breast cancer.⁴² In reference to the previous studies, there are 70% cumulative risks for BRCA carriers (BRCA1 & BRCA2) who are diagnosed with primary breast cancer. BRCA-related tumors frequently show different histopathological features that are incompetently differentiated but also highly proliferative.⁴³ Partial BRCA1 protein could be produced by a mutation in exon 11 that is encoded by the known exon 11 splice variant and it features a different function from whole BRCA1 protein.⁴²

Liquid Biopsy Biomarker

There are several studies that initiates on the capability of liquid biopsy to confirm the genomic profile, monitor responses of therapy, and evaluate the emergence of Development of Imaging and Liquid Biomarker Analysis for Breast Cancer Screening: A Review

Table 2. Primary protein biomarkers for breast cancer.

Protein	Description and Function			
Estrogen and progesterone receptor (hormone receptor)	Estrogen and Progesterone Receptor may encourage cell proliferation, tumor invasion and survival. Estrogen plays an important role for translocation by binding to its receptor into the nucleus as a transduction signal and as a transcription factor for several physiological responses in many target organs.			
HER2 (human epidermal growth factor Receptor)	High level expressions of mRNA and protein product by HER2 genes amplification could conduct sel sufficient and oncogenic resultant signaling in growth signals, continuous angiogenesis, high proliferatio rates, amplify metastasis processes and invasion that could encourage carcinogenesis.			
MIB1/Ki-67	Mib1/Ki-67 is a biomarker that is measured by proliferation index as parameters for predictive an prognostic marker. Although it is widely used, the ASCO guidelines have not included Mib1/Ki67 index a primary assessed marker for breast cancer prognosis due to the lack of standardization.			

resistance from patients.⁴⁴ In addition from the blood, there are several other body fluids like urine,⁴⁵ saliva,⁴⁶ cerebrospinal fluid,⁴⁷ pleural effusions,⁴⁸ and stool.⁴⁹ Serum or plasma that are utilized as biomarkers samples are potential for breast cancer screening as they accommodate valuable cellular and molecular content in the blood, which provide data about individual health information and could develop a great noninvasive diagnostics for breast cancer.⁵⁰

Some protein and peptide profiling in biological fluids has already become an interesting novel biomarkers for cancer patients.⁵¹ This method utilizes mass spectrometry (MS) as a tool to differentiate proteomic scheme of healthy individuals as controls and cancer patients.⁵² The earlystage of breast cancer detection emerged due to the increase of abnormality of total biomarkers from breast cancer patients that were up or down-regulated in comparison with healthy controls population.⁵³

Blood-based diagnostic assay

There are blood-borne tumor biomarkers have been introduced as a diagnostic assay to evaluate malignancy prior to the clinical diagnosis, such as the human epidermal growth factor receptor (HER2), carcinoembryonic antigen (CEA),⁵⁴ the oncogenic protein RS/DJ-1, and circulating cytokeratin fragments.⁵⁵

Blood-based Test using Multiple Reaction Monitoring

There is a blood-based test utilizing multiple reaction monitoring (MRM) as the method and measured by mass spectrometry that quantifies as 3 peptides: apolipoprotein C-1 (APOC1), carbonic anhydrase 1 (CAH1), and neural cell adhesion molecule L1-like protein (NCHL1) that illustrate different concentration level between healthy individuals as a control and breast cancer patients.56 APOC1 plays a vital role in lipoprotein metabolism that binds to fatty acids57 and reduces the addition of estrogen in cells. The amount is 0.7 times less in stage 1 breast cancer patients than the condition of healthy women.58 CAH1 enzymes are overexpressed and they increase rapidly through angiogenesis which is a key mechanism for tumors to develop to cancer.⁵⁹ The amount is 1.61 times higher for stage 1 breast cancer patients than the condition of healthy women.58 NCHL1 is closely related to cancer expression and metastasis⁶⁰ and the amount is 1.4 higher

more for stage 1 breast cancer patients than the condition of healthy women. $^{\rm 58}$

Blood-based Test for Detecting Copper (Cu)

The expression level of ATOX1 for patients may become a breast cancer biomarker for early stages diagnostics. Antioxidant 1 copper chaperone (ATOX1) plays an important role in cell migration process which is a core phase in metastasis.⁶¹ Copper (Cu) is one of the constituents of many enzymes that is required for several mechanisms in cancer such as angiogenesis, metastasis and proliferative immortality.^{62,63} The Cu concentration levels have been increased for breast cancers patients that have already developed a distant metastasis.⁶

Blood-based Test for Biomarker Fragments (ctDNA, CTCs, EXOs, MiRNA)

Novel approaches on the development of breast cancer diagnosis must provide potential biomarkers that contain relevant clinical information, meets the requirement, and less invasive methods (Figure 2). There are fragments in liquid biopsy samples such as circulating tumor DNA (ctDNA), circulating tumor cells (CTCs) or exosomes (EXOs) that are detached from tumorous cells (necrotic or apoptotic).⁴⁴ Low concentration of cell free DNA detected in the healthy person's blood and the quantity could be increased in breast cancer patient.⁶⁴

Cell free DNA (cfDNA) indicates the total DNA fragments in the blood samples that develop from three different sources which actively produce DNA: necrotic cells, apoptotic cells, and viable cells.⁶⁵ The main purpose of cfDNA-based analysis is to gather information towards the genetic changes in DNA fragments that are obtained from cancer cells such as circulating tumor DNA (ctDNA),⁶⁶ which could be analyzed by sequencing or digital polymerase chain reaction (dPCR).⁶⁷ There are some disadvantages from analyzing ctDNA which are the fraction of ctDNA is relatively low from the total cfDNA in cancer patients,68 only applicable for minority patients, as well as more expensive.⁶⁹ The amount fractions of ctDNA are ranging from 0 (undetectable) to 11.7%. There is no economical way to evaluate the amount of ctDNA fraction within the total of cfDNA⁶⁶ and still requires evidence in clinical trial.70

There are tumor cells called circulating tumor cells

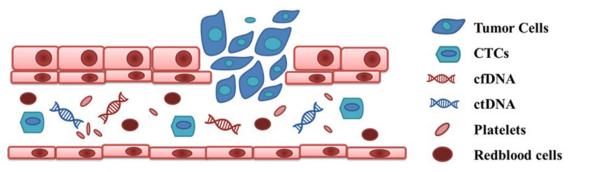


Figure 2. Biomarker fragments of breast cancer patient in blood vessel.

(CTCs) that possibly have been passively released from the major tumor and metastatic lesions into the bloodstream.⁴⁴ CTCs could be measured from blood samples of cancer patient and the total CTCs are correlated to overall survival and treatment outcomes.⁷¹ The concentration levels of CTCs that were detected in blood samples are relatively low, however it varies on the tumor types.⁷² CTC detection methods usually consist of several steps such as primary enrichment (due to low concentration), and cell isolation from blood cells with epithelial markers.⁷³ There are two common kits for detecting CTCs, CellSearch CTC test and AdnaTest.⁷⁴

Blood cells, smooth-muscle cells, platelets, endothelial cells, and immunocytes are known to be able to release exosomes which have significant roles in switching molecular information among cells.⁷⁵ They have already indicated that they contain proteins as well as some nucleic acids such as deoxyribonucleic acid (DNA), messenger RNA (mRNAs), and micro RNA (miRNAs). It is also shown that they can arrange the action of recipient cells. Exosomes could possibly be utilized as biomarkers of cancer.⁴⁴ Furthermore, exosomal miRNAs could be handful in cancer development as they could encourage angiogenesis and stimulate metastasis.⁷⁶

MicroRNAs are endogenous RNA molecules that contain between 19-25 nucleotides and have essential roles in post-transcriptional level as gene regulatory.77 MicroRNAs could develop combination with protein-coding genes messenger RNAs (mRNAs) and would lead to mRNA translational degradation or repression.78 MicroRNAs have a vital role in many cellular processes like differentiation, apoptosis, and proliferation. MicroRNAs alteration could also bring harmful transformation.⁷⁹ Several circulating miRNAs are diversely produced in the serum or plasma and could become potential biomarkers for breast cancer as the amount is different for healthy individuals and breast cancer patients. According to several studies, the most consistently upregulated miRNA is miR-21 as it functions as oncogene, while the most downregulated is miR-145 as it functions as tumor suppressor.⁸⁰

Salivary-based diagnostic assay

Biomarker research is constantly developing to the point where saliva is introduced as a very good diagnostic sample through technological advancements that could be collected non-invasively, simple, and could be gathered regularly without bringing discomfort to the subject.⁸¹ Salivary biomarkers analysis provide additional advantages such as monitoring clinical condition status and predicting diseases,⁸² but it requires combinatorial analysis of the biomarker profile to achieve appropriate level of specificity and sensitivity.²⁸

Vascular endothelial growth factor (VEGF), Carcinoembryonic antigen (CEA), and Epidermal growth factor (EGF)

Breast cancer metastasis and tumor growth could be elevated by angiogenesis mechanism and some angiogenic components such as vascular endothelial growth factor (VEGF), carcinoembryonic antigen (CEA), and epidermal growth factor (EGF). They detected through enzymelinked immunosorbent assay (ELISA) and could be found in saliva samples of breast cancer patients.⁸³ The level of those biomarkers are increased in the saliva samples of breast cancer patients in comparison with healthy individuals, mostly when those biomarkers were analyzed together as a combination.²⁸

Autoantibodies - Mucin1 (MUC1), Human Epidermal Growth Factor Receptor (HER2)

There is a high interest of exploration in autoantibodies against tumor biomarkers that could be evaluated in saliva.⁸⁴ Autoantibodies that are expressed against tumor biomarkers could offer a beneficial approach such as providing noninvasive method for breast cancer diagnostics.28 Mucin1 (MUC1) is a transmembrane glycoprotein that is overexpressed by around 90% in breast tumor and it performs a crucial part in development of the cancer. When MUC1 is overexpressed, it would stimulate growth of cells, resistance of therapy, and metastasis in cancer.85 Human epidermal growth factor receptor (HER2) is one of the biomarkers that is already detected and overexpressed in breast cancer patient, so it could induce cell migration and potentially become a metastatic factor.86 Autoantibodies against MUC1 and HER2 have already been investigated by using immunoglobulins (IgM and IgG) and has been detected by enzyme-linked immunosorbent assay (ELISA) test. The immunoglobulins were remarkably higher in breast cancer patients than in healthy individuals.87

Development of Imaging and Liquid Biomarker Analysis for Breast Cancer Screening: A Review

Sialic acid

Sialic acid is biologically notable for glycoconjugates and could be altered in cancer patients, thus the sialylation processes of cell surface glycoconjugates are increased and could cause malignant cancer progression.⁸⁸ Sialic acid concentration of salivary samples in breast cancer patients have been significantly increased compared to healthy individuals.⁸⁹ From the result, it could be concluded that sialic acid establishes clinical importance as a diagnostic marker.²⁸

Metabolites - Proline, Valine

Metabolites are one of the biomarker classes that is widely discovered in saliva samples of breast cancer patients utilized for diagnostic purposes.²⁸ Some studies showed significant changes in amino acid (basic metabolites) profile of breast cancer patients such as proline and valine. The analytical techniques that are utilized for detecting metabolites are Gas chromatography–Mass spectrometry (GC-MS), Liquid chromatography–Mass spectrometry (LC-MS), and Nuclear magnetic resonance (NMR spectroscopy).⁹⁰

Problem and Risk for Breast Cancer Screening

Although breast cancer screening has several benefits for women, it also hosts potential harms such as the side effect of screening. Balancing between the advantages and the harms of breast cancer screening may be rather complicated due to several considerations such as establishing harms possibilities, deciding the prime ages to start regular screening, determining the best intervals of screening test, using the relevant multiple imaging methods, and preferences of women concerning to screening.⁹¹ Breast cancer screening illustrated some risks such as falsepositive diagnostic results,⁹² anxiety,⁹³ radiation exposure,⁹⁴ pain during procedures,⁹⁵ and overdiagnosis.⁹⁶

Diagnostic Kit Models for Breast Cancer Screening

Reliable diagnostic test is essential for breast cancer classification and biomarkers such as human epidermal growth factor receptor 2 (HER2), estrogen receptor (ER), and progesterone receptor (PR).⁹⁷ The best analysis for the molecular research should be specific, reliable, sensitive, and easy to perform.⁹⁸ There are some diagnostic kit models that have already been used for breast cancer screening such as NanoString nCounter[®], MammaTyper[®], CellSearch System[™], and AdnaTest BreastCancer[™].

NanoString nCounter[®]

NanoString nCounter[®] gene expression system is a digital quantification technology based on RNA that performs color-coded multiplexed target molecule. It establishes the transcripts counts of mRNA from a limited quantity of total RNA without any amplification.⁹⁹ NanoString nCounter[®] gene expression system was performed for quantizing mRNA expression level of human epidermal growth factor receptor 2 (HER2), estrogen receptor (ER), and progester-one receptor (PR).⁹⁸

MammaTyper[®]

MammaTyper[®] is a diagnostic test that performs quantification for messenger RNA (mRNA) expression of biomarker genes such as PGR (PR), MKI67 (Ki-67), ERBB2 (HER2), and ESR1 (ER) by real-time quantitative polymerase chain reaction (RT-qPCR). MammaTyper[®] also classifies the results into different molecular subtypes i.e. HER2 positive (non-luminal), triple negative (ductal), Luminal A-like, and Luminal B-like (HER2 positive/HER2 negative).¹⁰⁰

CellSearch System[™] and AdnaTest BreastCancer[™]

Both CellSearch System[™] and AdnaTest BreastCancer[™] are diagnostic tests for detecting circulating tumor cells (CTCs) in blood samples of breast cancer patients.¹⁰¹ Those methods contain the cell-enrichment step and the detection step. Mainly, the cell-enrichment step requires antibody-based magnetic capture towards to the epithelial cell adhesion molecule-1 (EpCAM) as a target,¹⁰² and could be detected using immunofluorescence for CellSearch System[™] and by measuring tumor-associated transcript (MUC-1, HER2, and GA733-2) with reverse transcriptase-polymerase chain reaction (RT-PCR) for AdnaTest[™] BreatsCancer.¹⁰³

Conclusion

Early-stage detection of breast cancer may significantly minimize death rates in breast cancer patients for longterm. The realization of early diagnostics and screening programs are fundamental principles of cancer prevention. This paper summarized the screening methods and kind of biomarkers which are frequently available for diagnosing early-stage breast cancer. The recent development of breast cancer screening that utilizes different models and methods such as biomarkers were being reviewed. The development of breast cancer models should be more sensitive, reliable, approachable and less harmful.

Author Contributions

AS: Drafting the work, TR: The conception and design of the work and revising the work, RA: The conception and design of the work, and revising the work. All the authors agreed to the published version of the manuscript.

Acknowledgements

Thanks to all members of the present study group for their ideas, suggestions, and support. This study is part of thesis proposal components for a master's degree at the Faculty of Pharmacy, Padjadjaran University, Sumedang, Indonesia.

Conflict of Interest

All authors here claim no involvement in a conflict of interest, financial or otherwise.

References

1. Momenimovahed Z, Salehiniya H. Epidemiological characteristics of and risk factors for breast cancer

in the world. Breast Cancer (Dove Med Press). 2019;11:151-64. doi:10.2147/bctt.s176070

- Tungsukruthai S, Petpiroon N, Chanvorachote P. Molecular mechanisms of breast cancer metastasis and potential anti-metastatic compounds. Anticancer Res. 2018;38(5):2607-18. doi:10.21873/anticanres.12502
- Stastny I, Dankova Z, Kajo K, Kubatka P, Golubnitschaja O, Zubor P. Aberrantly methylated cfDNA in body fluids as a promising diagnostic tool for early detection of breast cancer. Clin Breast Cancer. 2020;20(6):711-22. doi:10.1016/j.clbc.2020.05.009
- 4. Wang L. Early diagnosis of breast cancer. Sensors (Basel). 2017;17(7):1572. doi:10.3390/s17071572
- Greenwald ZR, El-Zein M, Bouten S, Ensha H, Vazquez FL, Franco EL. Mobile screening units for the early detection of cancer: A systematic review. Cancer Epidemiol Biomarkers Prev. 2017;26(12):1679-94. doi:10.1158/1055-9965.epi-17-0454
- Baltzer PAT, Kapetas P, Marino MA, Clauser P. New diagnostic tools for breast cancer. Memo. 2017;10(3):175-80. doi:10.1007/s12254-017-0341-5
- Kwon EJ. Synthetic Biomarkers for Cancer Detection and Diagnosis. In: National Academy of Engineering. Frontiers of Engineering: Reports on Leading-Edge Engineering from the 2018 Symposium. Washington (DC): National Academies Press (US); 2019. doi:10.17226/25333
- Senkus E, Kyriakides S, Ohno S, Penault-Llorca F, Poortmans P, Rutgers E, et al. Primary breast cancer: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. Ann Oncol. 2015;26(5):8-30. doi:10.1093/annonc/mdv298
- Coleman C. Early detection and screening for breast cancer. Semin Oncol Nurs. 2017;33(2):141-55. doi:10.1016/j.soncn.2017.02.009
- Fuller MS, Lee CI, Elmore JG. Breast cancer screening: an evidence-based update. Med Clin North Am. 2015;99(3):451-68. doi:10.1016/j.mcna.2015.01.002
- Wöckel A, Albert US, Janni W, Scharl A, Kreienberg R, Stüber T. The Screening, Diagnosis, Treatment, and Follow-Up of Breast Cancer. Dtsch Arztebl Int. 2018;115(18):316-23. doi:10.3238/arztebl.2018.0316
- McDonald ES, Clark AS, Tchou J, Zhang P, Freedman GM. Clinical diagnosis and management of breast cancer. J Nucl Med. 2016;57(Suppl 1):S9-S16. doi:10.2967/jnumed.115.157834
- Zeeshan M, Salam B, Khalid QSB, Alam S, Sayani R. Diagnostic accuracy of digital mammography in the detection of breast cancer. Cureus. 2018;10(4):e2448. doi:10.7759/cureus.2448
- Cheuk IWY, Shin VY, Kwong A. Detection of methylated circulating DNA as noninvasive biomarkers for breast cancer diagnosis. J Breast Cancer. 2017;20(1):12-9. doi:10.4048/jbc.2017.20.1.12
- 15. Skaane P. Breast cancer screening with digital breast tomosynthesis. Breast Cancer. 2016;24(1):32-41. doi:10.1007/s12282-016-0699-y

- 16. Ohashi R, Nagao M, Nakamura I, Okamoto T, Sakai S. Improvement in diagnostic performance of breast cancer: comparison between conventional digital mammography alone and conventional mammography plus digital breast tomosynthesis. Breast Cancer. 2018;25(5):590-6. doi:10.1007/s12282-018-0859-3
- 17. Tagliafico AS, Bignotti B, Rossi F, Signori A, Sormani MP, Valdora F, et al. Diagnostic performance of contrast-enhanced spectral mammography: Systematic review and meta-analysis. Breast. 2016;28:13-9. doi:10.1016/j.breast.2016.04.008
- Lewis TC, Pizzitola VJ, Giurescu ME, Eversman WG, Lorans R, Robinson KA, et al. Contrast-enhanced digital mammography: A single-institution experience of the first 208 cases. Breast J. 2016;23(1):67-76. doi:10.1111/tbj.12681
- Hodgson R, Heywang-Köbrunner SH, Harvey SC, Edwards M, Shaikh J, Arber M, et al. Systematic review of 3D mammography for breast cancer screening. Breast. 2016;27:52-61. doi:10.1016/j.breast. 2016.01.002
- 20. Powell JL, Hawley JR, Lipari AM, Yildiz VO, Erdal BS, Carkaci S. Impact of the addition of digital breast tomosynthesis (DBT) to standard 2d digital screening mammography on the rates of patient recall, cancer detection, and recommendations for short-term follow-up. Acad Radiol. 2016;24(3):302-7. doi:10.1016/j.acra.2016.10.001
- Schneble EJ, Graham LJ, Shupe MP, Flynt FL, Banks KP, Kirkpatrick AD, et al. Future directions for the early detection of recurrent breast cancer. J Cancer. 2014;5(4):291-300. doi:10.7150/jca.8017
- 22. Watkins EJ. Overview of breast cancer. JAAPA. 2019;32(10):13-7. doi:10.1097/01.jaa.0000580524.950 733.3d
- 23. Gareth ED, Nisha K, Yit L, Soujanye G, Emma H, Stephen D. MRI breast screening in high-risk women: cancer detection and survival analysis. Breast Cancer Res Treat. 2014;145(3):663-72. doi:10.1007/s10549-014-2931-9
- 24. Ohuchi N, Suzuki A, Sobue T, Kawai M, Yamamoto S, Zheng YF, et al. Sensitivity and specificity of mammography and adjunctive ultrasonography to screen for breast cancer in the Japan Strategic Anticancer Randomized Trial (J-START): a randomised controlled trial. Lancet. 2016;387(10016):341-8. doi:10.1016/s0140-6736(15)00774-6
- 25. Scheel JR, Lee JM, Sprague BL, Lee CI, Lehman CD. Screening ultrasound as an adjunct to mammography in women with mammographically dense breasts. Am J Obstet Gynecol. 2015;212(1):9-17. doi:10.1016/j. ajog.2014.06.048
- 26. Hooley RJ, Scoutt LM, Philpotts LE. Breast ultrasonography: State of the art. Radiology. 2013;268(3):642-59. doi:10.1148/radiol.13121606
- 27. Mittal S, Kaur H, Gautam N, Mantha AK. Biosensors

for breast cancer diagnosis: A review of bioreceptors, biotransducers and signal amplification strategies. Biosens Bioelectron. 2017;88:217-31. doi:10.1016/j. bios.2016.08.028

- Porto-Mascarenhas EC, Assad DX, Chardin H, Gozal D, De Luca Canto G, Acevedo AC, et al. Salivary biomarkers in the diagnosis of breast cancer: A review. Crit Rev Oncol Hematol. 2017;110:62-73. doi:10.1016/j.critrevonc.2016.12.009
- 29. Mishra A, Verma M. Cancer biomarkers: Are we ready for the prime time. Cancers (Basel). 2010;2(1):190-208. doi:10.3390/cancers2010190
- Diaconu I, Cristea C, Hârceagă V, Marrazza G, Berindan-Neagoe I, Săndulescu R. Electrochemical immunosensors in breast and ovarian cancer. Clin Chim Acta. 2013;425:128-38. doi:10.1016/j. cca.2013.07.017
- 31. You K, Park S, Ryu JM, Kim I, Lee SK, Yu J, et al. Comparison of core needle biopsy and determining surgical specimens in intrinsic biological subtypes of breast cancer with immunohistochemistry. J Breast Cancer. 2017;20(3):297-303. doi:10.4048/jbc.2017.20.3.297
- 32. Fragomeni SM, Sciallis A, Jeruss, JS. Molecular subtypes and local-regional control of breast cancer. Surg Oncol Clin N Am. 2018;27(1):95-120. doi:10.1016/j.soc.2017.08.005
- Bertozzi S, Londero AP, Seriau L, Vora RD, Cedolini C, Mariuzzi L. Biomarkers in breast cancer. In: Biomarker -Indicator of Abnormal Physiological Process, Begum G. editor. London: IntechOpen; 2018. doi:10.5772/ intechopen.77320
- Tsang JYS, Tse GM. Molecular classification of breast cancer. Advances Adv Anat Pathol. 2020;27(1):27-35. doi:10.1097/pap.00000000000232
- 35. Renoir JM, Marsaud V, Lazennec G. Estrogen receptor signaling as a target for novel breast cancer therapeutics. Biochem Pharmaco. 2013;85(4):449-65. doi:10.1016/j.bcp.2012.10.018
- 36. Grober OM, Mutarelli M, Giurato G, Ravo M, Cicatiello L, De Filippo MR, et al. Global analysis of estrogen receptor beta binding to breast cancer cell genome reveals an extensive interplay with estrogen receptor alpha for target gene regulation. BMC Genomics. 2011;12:36. doi:10.1186/1471-2164-12-36
- 37. Nofech-Mozes S, Vella ET, Dhesy-Thind S, Hagerty KL, Mangu PB, Temin S, et al. Systematic Review on Hormone Receptor Testing in Breast Cancer. Appl Immunohistochem Mol Morphol. 2012;20(3):214-63. doi:10.1097/pai.0b013e318234aa 12
- 38. Hammond MEH, Hayes DF, Dowsett M, Allred DC, Hagerty KL, Badve S, et al. American Society of Clinical Oncology/College of American Pathologists guideline recommendations for immunohistochemical testing of estrogen and progesterone receptors in breast cancer. Arch Pathol Lab Med. 2010;134(7):e48-72.

doi:10.5858/134.7.e48

- 39. Wolff AC, Hammond MEH, Hicks DG, Dowsett M, McShane LM, Allison KH, et al. Recommendations for human epidermal growth factor receptor 2 testing in breast cancer: American Society of Clinical Oncology/College of American Pathologists clinical practice guideline update. Arch Pathol Lab Med. 2014;138(2):241-56. doi:10.5858/arpa.2013-0953-sa
- 40. Wolff AC, Hammond MEH, Schwartz JN, Hagerty KL, Allred DC, Cote RJ, et al. American society of clinical oncology/college of american pathologist's guideline recommendations for human epidermal growth factor receptor 2 testing in breast cancer. Arch Pathol Lab Med. 2007;131(1):18-43. doi:10.5858/2007-131-18-ASOCCO
- 41. Xiang H, Xin L, Liu Q, Zhang H, Zhang S, Ye J, et al. Clinicopathological analysis of early-stage breast cancer patients that meet indications for BRCA1/2 genetic testing. Chin J Cancer Res. 2020;32(2):163-74. doi:10.21147/j.issn.1000-9604.2020.02.04
- 42. Rebbeck TR, Mitra N, Wan F, Sinilnikova OM, Healey S, McGuffog L, et al. Association of type and location of BRCA1 and BRCA2 mutations with risk of breast and ovarian cancer. JAMA. 2015;313(13):1347. doi:10.1001/jama.2014.5985
- 43. Fountzilas E, Konstantopoulou I, Vagena A, Apostolou P, Papadimitriou C, Christodoulou C, et al. Pathology of BRCA1- and BRCA2- associated breast cancers: Known and less known connections. Clin Breast Cancer. 2020;20(2):152-9. doi:10.1016/j. clbc.2019.08.003
- 44. Siravegna G, Marsoni S, Siena S, Bardelli A. Integrating liquid biopsies into the management of cancer. Nat Rev Clin Oncol. 2017;14(9):531-48. doi:10.1038/ nrclinonc.2017.14
- 45. Reckamp KL, Melnikova VO, Karlovich C, Sequist LV, Camidge DR, Wakelee H, et al. A Highly sensitive and quantitative test platform for detection of NSCLC EGFR mutations in urine and plasma. J Thorac Oncol. 2016;11(10):1690-700. doi:10.1016/j.jtho.2016.05.035
- 46. Wang Y, Springer S, Mulvey CL, Silliman N, Schaefer J, Sausen M, et al. Detection of somatic mutations and HPV in the saliva and plasma of patients with head and neck squamous cell carcinomas. Sci Transl Med. 2015;7(293):293ra104. doi:10.1126/scitranslmed.aaa 8507
- 47. De Mattos-Arruda L, Mayor R, Ng CKY, Weigelt B, Martínez-Ricarte F, Torrejon D, et al. Cerebrospinal fluid-derived circulating tumour DNA better represents the genomic alterations of brain tumours than plasma. Nature Communications. 2015;6:8839. doi:10.1038/ncomms9839
- Kimura H, Fujiwara Y, Sone T, Kunitoh H, Tamura T, Kasahara K, et al. EGFR mutation status in tumourderived DNA from pleural effusion fluid is a practical basis for predicting the response to gefitinib. Br J Cancer. 2006;95(10):1390-5. doi:10.1038/sj.bjc.

6603428

- Diehl F, Schmidt K, Durkee KH, Moore KJ, Goodman SN, Shuber AP, et al. Analysis of mutations in DNA Isolated from plasma and stool of colorectal cancer patients. Gastroenterology. 2008;135(2):489-98. doi:10.1053/j.gastro.2008.05.039
- Núñez C. Blood-based protein biomarkers in breast cancer. Clin Chim Acta. 2019;490:113-27. doi:10.1016/j.cca.2018.12.028.
- Tsai TH, Song E, Zhu R, Di Poto C, Wang M, Luo Y, et al. LC-MS/MS-based serum proteomics for identification of candidate biomarkers for hepatocellular carcinoma. Proteomics. 2015;15(13):2369-81. doi:10.1002/pmic. 201400364
- 52. Fan NJ, Kang R, Ge XY, Li M, Liu Y, Chen HM, et al. Identification alpha-2-HS-glycoprotein precursor and tubulin beta chain as serology diagnosis biomarker of colorectal cancer. Diagn Pathol. 2014;9:53. doi:10.1186/1746-1596-9-53
- 53. Callesen AK, Madsen JS, Vach W, Kruse TA, Mogensen O, Jensen ON. Serum protein profiling by solid phase extraction and mass spectrometry: A future diagnostics tool. Proteomics. 2009;9(6):1428-41. doi:10.1002/pmic.200800382
- 54. Di Gioia D, Dresse M, Mayr D, Nagel D, Heinemann V, Stieber P. Serum HER2 in combination with CA 15-3 as a parameter for prognosis in patients with early breast cancer. Clin Chim Acta. 2015;440:16-22. doi:10.1016/j.cca.2014.11.001
- 55. Harris L, Fritsche H, Mennel R, Norton L, Ravdin P, Taube S, et al. American society of clinical oncology 2007 update of recommendations for the use of tumor markers in breast cancer. J Clin Oncol. 2007;25(33):5287-312. doi:10.1200/JCO.2007.14.2364
- 56. Lee HB, Kang UB, Moon HG, Lee J, Lee KM, Yi M, et al. Development and validation of a novel plasma protein signature for breast cancer diagnosis by using multiple reaction monitoring-based mass spectrometry. Anticancer Res. 2015;35(11):6271-9.
- 57. Sun Y, Zhang J, Guo F, Zhao W, Zhan Y, Liu C, et al. Identification of apolipoprotein C-I peptides as a potential biomarker and its biological roles in breast cancer. Med Sci Monit. 2016;22:1152-60. doi:10.12659/ msm.896531
- 58. Kim Y, Kang UB, Kim S, Lee HB, Moon HG, Han W, et al. A Validation Study of a Multiple Reaction Monitoring-Based Proteomic Assay to Diagnose Breast Cancer. J Breast Cancer. 2019;22(4):579-86. doi:10.4048/jbc.2019.22.e57
- Mboge M, Mahon B, McKenna R, Frost S. Carbonic anhydrases: Role in pH control and cancer. Metabolites. 2018;8(1):19. doi:10.3390/metabo8010019
- 60. Wu JD, Hong CQ, Huang WH, Wei XL, Zhang F, Zhuang YX, et al. L1 cell adhesion molecule and its soluble form sl1 exhibit poor prognosis in primary breast cancer patients. Clin Breast Cancer. 2018;18(5):851-61. doi:10.1016/j.clbc.2017.12.011

- 61. Blockhuys S, Brady DC, Wittung-Stafshede P. Evaluation of copper chaperone ATOX1 as prognostic biomarker in breast cancer. Breast Cancer. 2020;27(3):505-9. doi:10.1007/s12282-019-01044-4
- Denoyer D, Masaldan S, La Fontaine S, Cater MA. Targeting copper in cancer therapy: 'Copper That Cancer'. Metallomics. 2015;7(11):1459-76. doi:10.1039/C5MT00149H
- 63. Choi R, Kim MJ, Sohn I, Kim S, Kim I, Ryu JM, et al. Serum trace elements and their associations with breast cancer subgroups in korean breast cancer patients. Nutrients. 2018;11(1):37. doi:10.3390/ nu11010037
- 64. Panagopoulou M, Karaglani M, Balgkouranidou I, Biziota E, Koukaki T, Karamitrousis E, et al. Circulating cell-free DNA in breast cancer: size profiling, levels, and methylation patterns lead to prognostic and predictive classifiers. Oncogene. 2019;38:3387-401. doi:10.1038/s41388-018-0660-y
- 65. Jahr S, Hentze H, Englisch S, Hardt D, Fackelmayer FO, Hesch RD, et al. DNA fragments in the blood plasma of cancer patients: quantitations and evidence for their origin from apoptotic and necrotic cells. Cancer Res. 2001;61(4):1659-65.
- 66. Lu JL, Liang ZY. Circulating free DNA in the era of precision oncology: Pre- and post-analytical concerns. Chronic Dis Transl Med. 2016;2(4):223-30. doi:10.1016/j.cdtm.2016.12.001
- 67. Li S, Lai H, Liu J, Liu Y, Jin L, Li Y, et al. Circulating tumor DNA predicts the response and prognosis in patients with early breast cancer receiving neoadjuvant chemotherapy. JCO Precision Oncol. 2020;4:244-57. doi:10.1200/PO.19.00292
- Wurdinger T, In'tVeld SGJG, Best MG. Platelet RNA as pan-tumor biomarker for cancer detection. Cancer Res. 2020;80(7):1371-3. doi:10.1158/0008-5472.can-19-3684.
- 69. Newman AM, Bratman SV, To J, Wynne JF, Eclov NCW, Modlin LA, et al. An ultrasensitive method for quantitating circulating tumor DNA with broad patient coverage. Nat Med. 2014;20(5):548-54. doi:10.1038/nm.3519
- Merker JD, Oxnard GR, Compton C, Diehn M, Hurley P, Lazar AJ, et al. Circulating tumor DNA analysis in patients with cancer: American society of clinical oncology and college of american pathologists joint review. J Clin Oncol. 2018;36(16):1631-41. doi:10.1200/JCO.2017.76.8671
- 71. Krebs MG, Metcalf RL, Carter L, Brady G, Blackhall FH, Dive C. Molecular analysis of circulating tumour cells—biology and biomarkers. Nat Rev Clin Oncol. 2014;11(3):129-44. doi:10.1038/nrclinonc.2013.253
- 72. Haber DA, Velculescu VE. Blood-based analyses of cancer: Circulating tumor cells and circulating tumor DNA. Cancer Discov. 2014;4(6):650-61. doi:10.1158/2159-8290.cd-13-1
- 73. Thery L, Meddis A, Cabel L, Proudhon C, Latouche

A, Pierga JY, et al. Circulating tumor cells in early breast cancer. JNCI Cancer Spectr. 2019;3(2):pkz026. doi:10.1093/jncics/pkz026

- Eroglu Z, Fielder O, Somlo, G. Analysis of circulating tumor cells in breast cancer. J Natl Compr Canc Netw. 2013;11(8):977-85. doi:10.6004/jnccn.2013.0118
- 75. Liao J, Liu R, Yin L, Pu Y. Expression Profiling of Exosomal miRNAs Derived from Human Esophageal Cancer Cells by Solexa High-Throughput Sequencing. Int J Mol Sci. 2014;15(9):15530-51. doi:10.3390/ ijms150915530.
- 76. Zhang J, Li S, Li L, Li M, Guo C, Yao J, et al. Exosome and exosomal microRNA: Trafficking, sorting, and function. Genomics Proteomics Bioinformatics. 2015;13(1):17-24. doi:10.1016/j.gpb.2015.02.001
- 77. Bertoli G, Cava C, Castiglioni I. MicroRNAs: New biomarkers for diagnosis, prognosis, therapy prediction and therapeutic tools for breast cancer. Theranostics. 2015;5(10):1122-43. doi:10.7150/thno. 11543
- Matamala N, Vargas MT, Gonzalez-Campora R, Minambres R, Arias JI, Menendez P, et al. Tumor microRNA expression profiling identifies circulating microRNAs for early breast cancer detection. Clin Chem. 2015;61(8):1098-106. doi:10.1373/ clinchem.2015.238691
- Sayed D, Abdellatif M. MicroRNAs in development and disease. Physiol Rev. 2011;91(3):827-87. doi:10.1152/physrev.00006.2010
- Adhami M, Haghdoost AA, Sadeghi B, Malekpour AR. Candidate miRNAs in human breast cancer biomarkers: a systematic review. Breast Cancer. 2017;25(2):198-205. doi:10.1007/s12282-017-0814-8
- Streckfus C, Bigler L. Saliva as a diagnostic fluid. Oral Dis. 2002;8(2):69-76. doi:10.1034/j.1601-0825.2002.10834.x
- Sugimoto M, Wong DT, Hirayama A, Soga T, Tomita M. Capillary electrophoresis mass spectrometrybased saliva metabolomics identified oral, breast and pancreatic cancer-specific profiles. Metabolomics. 2009;6(1):78-95. doi:10.1007/s11306-009-0178-y
- Brooks M, Wang J, Li Y, Zhang R, Elashoff D, Wong D. Salivary protein factors are elevated in breast cancer patients. Mol Med Rep. 2008;1(3):375-8. doi:10.3892/ mmr.1.3.375
- 84. Arif S, Qudsia S, Urooj S, Chaudry N, Arshad A, Andleeb S. Blueprint of quartz crystal microbalance biosensor for early detection of breast cancer through salivary autoantibodies against ATP6AP1. Biosens Bioelectron. 2015;65:62-70. doi:10.1016/j. bios.2014.09.088
- Nath S, Mukherjee P. MUC1: a multifaceted oncoprotein with a key role in cancer progression. Trends Mol Med. 2014;20(6):332-42. doi:10.1016/j. molmed.2014.02.007
- 86. Johnson E, Seachrist DD, DeLeon-Rodriguez CM, Lozada KL, Miedler J, Abdul-Karim FW, et al. HER2/

ErbB2-induced breast cancer cell migration and invasion require p120 catenin activation of Rac1 and Cdc42. J Biol Chem. 2010;285(38):29491-501. doi:10.1074/jbc.m110.136770

- Laidi F, Bouziane A, Errachid A, Zaoui F. Usefulness of salivary and serum auto-antibodies against tumor biomarkers HER2 and MUC1 in breast cancer screening. Asian Pac J Cancer Prev. 2016;17(1):335-9. doi:10.7314/apjcp.2016.17.1.335
- Shah MH, Telang SD, Shah PM, Patel PS. Tissue and serum α2-3- and α2-6-linkage specific sialylation changes in oral carcinogenesis. Glycoconj J. 2008;25(3):279-90. doi:10.1007/s10719-007-9086-4
- Oztürk LK, Emekli-Alturfan E, Kaşikci E, Demir G, Yarat A. Salivary total sialic acid levels increase in breast cancer patients: a preliminary study. Med Chem. 2011;7(5):443-7. doi:10.2174/157340611796799230
- 90. Cheng F, Wang Z, Huang Y, Duan Y, Wang X. Investigation of salivary free amino acid profile for early diagnosis of breast cancer with ultra-performance liquid chromatography-mass spectrometry. Clinica Chimica Acta. 2015;447:23-31. doi:10.1016/j.cca. 2015.05.008
- 91. Nelson HD, Pappas M, Cantor A, Griffin J, Daeges M, Humphrey L. Harms of breast cancer screening: systematic review to update the 2009 U.S. Preventive services task force recommendation. Ann Intern Med. 2016;164(4):256-7. doi:10.7326/m15-0970
- 92. Hubbard RA, Kerlikowske K, Flowers CI, Yankaskas BC, Zhu W, Miglioretti DL. Cumulative probability of false-positive recall or biopsy recommendation after 10 Years of screening mammography. Ann Intern Med. 2011;155(8):481. doi:10.7326/0003-4819-155-8-201110180-00004
- 93. Keyzer-Dekker CMG, De Vries J, VanEsch L, Ernst MF, Nieuwenhuijzen GAP, Roukema JA, et al. Anxiety after an abnormal screening mammogram is a serious problem. Breast. 2012;21(1):83-8. doi:10.1016/j.breast. 2011.08.137
- 94. Yaffe MJ, Mainprize JG. Risk of radiation-induced breast cancer from mammographic screening. Radiology. 2011;258(1):98-105. doi:10.1148/radiol. 10100655
- 95. Whelehan P, Evans A, Wells M, MacGillivray S. The effect of mammography pain on repeat participation in breast cancer screening: A systematic review. Breast. 2013;22(4):389-94. doi:10.1016/j.breast.2013.03.003
- 96. Puliti D, Duffy SW, Miccinesi G, De Koning H, Lynge E, Zappa M, et al. Overdiagnosis in mammographic screening for breast cancer in europe: A literature review. J Med Screen. 2012;19(1):42-56. doi:10.1258/jms.2012.012082
- 97. Schnitt SJ. Classification and prognosis of invasive breast cancer: from morphology to molecular taxonomy. Mod Pathol. 2010;23(2):60-4. doi:10.1038/ modpathol.2010.33
- 98. Hyeon J, Cho SY, Hong ME, Kang SY, Do I, Im YH, et

al. NanoString nCounter[®] approach in breast cancer: A comparative analysis with quantitative real-time polymerase chain reaction, in situ hybridization, and immunohistochemistry. J Breast Cancer. 2017;20(3):286-96. doi:10.4048/jbc.2017.20.3.286

- 99. Geiss GK, Bumgarner RE, Birditt B, Dahl T, Dowidar N, Dunaway DL, et al. Direct multiplexed measurement of gene expression with color-coded probe pairs. Nature Biotechnol. 2008;26(3):317-25. doi:10.1038/nbt1385
- 100. Varga Z, Lebeau A, Bu H, Hartmann A, Penault-Llorca F, Guerini-Rocco E, et al. An international reproducibility study validating quantitative determination of ERBB2, ESR1, PGR, and MKI67 mRNA in breast cancer using MammaTyper[®]. Breast Cancer Res. 2017;19(1):55. doi:10.1186/s13058-017-0848-z
- 101. Müller V, Riethdorf S, Rack B, Janni W, Fasching PA,

Solomayer E, et al. Prognostic impact of circulating tumor cells assessed with the CellSearch System[™] and AdnaTest Breast[™] in metastatic breast cancer patients: the detect study. Breast Cancer Res. 2012;14(4):118. doi:10.1186/bcr3243

- 102. Osta WA, Chen Y, Mikhitarian K, Mitas M, Salem M, Hannun YA, et al. EpCAM Is overexpressed in breast cancer and is a potential target for breast cancer gene therapy. Cancer Res. 2004;64(16):5818-24. doi:10.1158/0008-5472.can-04-0754
- 103. Andreopoulou E, Yang LY, Rangel KM, Reuben JM, Hsu L, Krishnamurthy S, et al. Comparison of assay methods for detection of circulating tumor cells in metastatic breast cancer: AdnaGen AdnaTest BreastCancer Select/Detect[™] versus Veridex CellSearch[™] system. Int J Cancer. 2011;130(7):1590-7. doi:10.1002/ijc.26111